



The Brazilian Journal of INFECTIOUS DISEASES

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Original article

CTX-M-producing *Escherichia coli* and *Klebsiella pneumoniae* isolated from community-acquired urinary tract infections in Valledupar, Colombia

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ARTICLE INFO

Article history:

Received 11 February 2012

Accepted 26 May 2012

Available online 8 September 2012

Keywords:

ESBL

CTX-M

E. coli

K. pneumoniae

Community-acquired UTIs

Colombia

ABSTRACT

Objective: Describe the presence of CTX-M-1 phylogenetic subgroup extended-spectrum β -lactamases (ESBL), associated with TEM and SHV genes, and the gene encoding cephalosporinase, CMY-2 in *Escherichia coli* and *Klebsiella pneumoniae* isolates from community-acquired urinary tract infections.

Methods: 102 *E. coli* and 21 *K. pneumoniae* were collected from patients with culture-proven urinary tract infection (UTI), during February and March, 2011. Antimicrobial susceptibility test was performed by disk diffusion according to the standards of the Clinical Laboratory Standard Institute. Screening for cephalosporins-resistant *E. coli* and *K. pneumoniae* was performed by PCR assay for *bla*_{TEM}, *bla*_{SHV}, *bla*_{CTX-M-1,-2,-8,-9}, *bla*_{PER-2} and *bla*_{CMY-2} genes. Statistical analysis was performed by chi-squared test and multivariate logistic regression analysis.

Results: ESBL production was detected in 12 (11.7%) *E. coli* and four (19%) *K. pneumoniae* isolates. TEM ESBLs were detected in seven *E. coli* and three *K. pneumoniae* isolates. SHV ESBLs were found in four *K. pneumoniae* isolates. CTX-M-1 phylogenetic subgroup was positive in seven *E. coli* and three *K. pneumoniae* isolates. CMY-2 β -lactamase gene was detected in nine *E. coli* and one *K. pneumoniae* isolates. A significant association of ESBL expression in *E. coli* was observed with resistance to tobramycin ($p \leq 0.001$), tetracycline ($p = 0.043$), and ciprofloxacin ($p \leq 0.001$). In *K. pneumoniae* isolates, significant association was found with resistance to tobramycin and ciprofloxacin ($p = 0.006$), and trimethoprim-sulfamethoxazole ($p = 0.043$). Multivariate analyses did not show association between ESBL production in *E. coli* and *K. pneumoniae*, and resistance to non- β -lactams drugs.

Conclusions: CTX-M ESBL in uropathogens isolated from the community is cause for concern due to the enormous potential for multidrug resistance from strains that produce these enzymes, which could lead to failure of empirically-administered therapies and development of complicated UTIs.

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<http://dx.doi.org/10.1016/j.bjid.2012.05.001>

Introduction

Infections caused by extended-spectrum β -lactamases (ESBLs)-producing *E. coli* and *K. pneumoniae* have been described in the hospital setting,¹ although during the last decade these organisms have begun to disseminate into the community, becoming an emerging public health problem.² ESBLs are enzymes commonly associated with TEM and SHV. However, many *E. coli* and *K. pneumoniae* isolates possess the gene encoding *ampC* cephalosporinase.³ Moreover, CTX-M-type ESBLs have emerged within the community, particularly among *E. coli* and *K. pneumoniae* isolated from urinary tract infections (UTIs), with a widespread prevalence and multidrug resistance in many countries worldwide.^{2,4} Multidrug resistance expressed by CTX-M producing isolates from the community is often associated with the presence of multiple ESBLs genes, as well as aminoglycoside and quinolone resistance genes,^{5,6} thereby limiting the choice of effective antimicrobial drugs.⁷

The aim of this study was to describe the presence of the CTX-M-1 phylogenetic subgroup ESBL associated with TEM and SHV genes and the gene encoding cephalosporinase, CMY-2, in *E. coli* and *K. pneumoniae* isolates from community-acquired UTIs.

Materials and methods

Bacterial isolates

During a period of two months (February and March) in 2011, 123 isolates of enterobacteria (102 *Escherichia coli* and 21 *Klebsiella pneumoniae*), were collected from patients with culture-proven UTI evaluated in four clinical microbiology laboratories in Valledupar, Colombia. Cultures were obtained from midstream-collected urine. A uroculture was considered positive when it presented a bacterial count of more than 100,000 colony-forming units (CFU)/mL in a urine specimen collected midstream.⁸ *E. coli* and *K. pneumoniae* identification was performed according to conventional method.⁹ This study was based on laboratory records and not on medical records. Organisms were isolated from non-hospitalized patients during an outpatient visit to any of the four clinical microbiology laboratories.

This study did not require an approval from the Ethics Committee of Institute for Tropical Biological Research from Universidad de Córdoba, Colombia.

Susceptibility testing and ESBL detection

Antimicrobial susceptibility and interpretation were performed by disk diffusion according to Clinical Laboratory Standard Institute (CLSI).¹⁰ The antimicrobials used were: piperacillin-tazobactam (100 μ g/10g), ceftazidime (30 μ g), ceftriaxone (30 μ g), cefepime (30 μ g), aztreonam (30 μ g), cefoxitin (30 μ g), ertapenem (10 μ g), imipenem (10 μ g), tobramycin (10 μ g), ciprofloxacin (5 μ g), trimethoprim-sulfamethoxazole (25 μ g), and tetracycline (30 μ g) from Oxoid, Basingstoke, United Kingdom. ESBL phenotype was detected

by interpretation of the CLSI criteria and disk combination method.¹¹ *E. coli* ATCC® 25922 and *K. pneumoniae* ATCC® 700603 were used as control strains.

PCR amplification

Screening of cephalosporins-resistant *E. coli* and *K. pneumoniae* isolates was performed by PCR assay for *bla*_{TEM}, *bla*_{SHV}, *bla*_{CTX-M-1}, *bla*_{CTX-M-2}, *bla*_{CTX-M-8}, *bla*_{CTX-M-9}, *bla*_{PER-2} and *bla*_{CMY-2} genes using the primers described elsewhere.¹²⁻¹⁶

Statistical analysis

A statistical comparison of the frequencies of ESBL presence in *E. coli* and *K. pneumoniae* isolates was conducted by chi-squared test and multivariate logistic regression analysis. A value of $p < 0.05$ was considered significant. All statistical calculations were performed by using the Statistical Analysis System version 9.1.3 (SAS, Cary, North Carolina, USA).

Results

Antibiotic susceptibility

Susceptibility profile expressed by 123 enterobacteria isolates (102 *E. coli* and 21 *K. pneumoniae*) is shown in Table 1. *E. coli* isolates included in this study presented resistance to broad-spectrum cephalosporins in 23.8% of cases. In *K. pneumoniae* the resistance rate to broad-spectrum cephalosporins was 12.7%. Resistance expressed by *E. coli* and *K. pneumoniae* isolates was of substantial level to non- β -lactam agents (Table 1). None of the strains were resistant to carbapenems.

Characteristics of cephalosporins-resistant enterobacteria

Eighteen broad-spectrum cephalosporins-resistant isolates were tested by disk combination method, and ESBL production was found in 11 *E. coli* (10.7%) and four *K. pneumoniae* (19%) isolates by phenotypic confirmation test, as the zone of inhibition increased in presence of clavulanic acid. Three isolates showed full resistance to third-generation cephalosporins and to a combination of both cefotaxime and ceftazidime with clavulanic acid, a phenotype consistent with AmpC β -lactamase production (Table 2).

Molecular analysis by amplification of purified DNA in 18 cephalosporins-resistant isolates yielded a PCR product with specific primers for *bla*_{TEM}, *bla*_{SHV} and *bla*_{CTX-M-1} genes in 16 isolates. The molecular method found one additional ESBL-positive *E. coli* isolate that the phenotypic method had not detected (Table 2). Therefore, ESBL production was detected in 12 (11.7%) *E. coli* isolates; *bla*_{CTX-M-2}, *bla*_{CTX-M-8}, *bla*_{CTX-M-9}, and *bla*_{PER-2} were not detected by PCR screening. TEM ESBLs were detected in ten ESBL producers (seven *E. coli* and three *K. pneumoniae*). SHV ESBLs were found in four *K. pneumoniae* isolates. PCR amplification by CTX-M type ESBL was positive for CTX-M-1 phylogenetic subgroup in seven *E. coli* and three *K. pneumoniae*. CMY-2 β -lactamase gene was detected in nine *E. coli* and one *K. pneumoniae* isolates. A combination of

Table 1 – Percentage of antibiotic susceptibility expressed by *E. coli* and *K. pneumoniae* isolates.

Antimicrobial drug	Breakpoint (mm)	<i>E. coli</i>				<i>K. pneumoniae</i>			
		%R	%I	%S	%R 95% CI	%R	%I	%S	%R 95% CI
Ceftazidime	18-20	12.7	0	87.3	7.2-21.1	23.8	0	76.2	9.1-47.5
Ceftriaxone	20-22	12.7	0	87.3	7.2-21.1	23.8	0	76.2	9.1-47.5
Cefepime	15-17	12.7	0	87.3	7.2-21.1	23.8	0	23.8	9.1-47.5
Aztreonam	18-20	12.7	0	87.3	7.2-21.1	23.8	0	23.8	9.1-47.5
Ertapenem	20-22	0	0	100	0.0-4.5	0	0	100	0.3-25.9
Imipenem	20-22	0	0	100	0.0-4.5	0	0	100	0.0-19.2
Tobramycin	13-14	21.6	0	78.4	14.3-31.1	28.6	0	71.4	12.2-52.3
Amikacin	15-16	11.8	0	88.20%	6.5-20.1	14.3	0	85.7	3.8-37.4
Ciprofloxacin	16-20	38.2	0	61.8	28.9-48.4	33.3	0	66.7	15.5-56.9
SXT	11-15	50	0	50	40.0-60.0	23.8	0	76.2	9.1-47.5
Tetracycline	12-14	60.8	0	39.2	50.6-70.2	19	0	81	6.3-42.5

R, resistance; I, intermediate resistance; S, susceptible; SXT, trimethoprim-sulfamethoxazole; CI, confidence interval.

Table 2 – Molecular and phenotypic characteristics of third-generation cephalosporins-resistant *E. coli* and *K. pneumoniae* isolates.

Pathogen	No. isolates	Disk combination method	ESBL genes				Susceptibility by disk diffusion CLSI								
			<i>bla</i> _{TEM}	<i>bla</i> _{SHV}	<i>bla</i> _{CTX-M-1}	<i>bla</i> _{CMY-2}	AK	FOX	CIP	ETP	IPM	TZP	TCY	TOB	SXT
<i>E. coli</i>	35	+	+	-	+	+	S	S	R	S	S	S	R	R	S
	40	+	-	-	+	+	S	S	R	S	S	R	R	R	R
	102	+	+	-	-	+	S	R	R	S	S	S	S	S	S
	108	+	-	-	-	-	S	S	R	S	S	R	R	S	R
	525	-	-	-	-	+	R	R	R	S	S	R	R	R	R
	527	+	+	-	+	+	S	S	R	S	S	S	R	S	R
	551	+	+	-	+	+	S	S	R	S	S	S	R	R	R
	576	+	-	-	-	-	S	S	R	S	S	S	R	S	S
	580	-	-	-	+	+	S	R	R	S	S	R	R	R	R
	612	+	-	-	+	+	S	S	R	S	S	S	R	R	R
	673	+	+	-	-	-	S	S	R	S	S	R	R	R	R
	678	+	+	-	+	+	R	R	R	S	S	R	R	R	R
	698	+	+	-	-	+	S	S	R	S	S	S	R	R	R
<i>K. pneumoniae</i>	111	+	-	+	+	-	S	S	R	S	S	R	S	R	R
	587	+	+	+	+	-	S	S	R	S	S	S	R	R	R
	672	-	-	-	-	+	R	R	S	S	S	R	S	S	S
	710	+	+	+	+	-	R	S	R	S	S	R	S	R	R
	5125	+	+	+	-	+	R	R	R	S	S	R	S	R	S

AK, amikacin; FOX, ceftazidime; CIP, ciprofloxacin; ETP, ertapenem; IPM, imipenem; TZP, piperacillin/ tazobactam; TCY, tetracycline; TOB, tobramycin; SXT, trimethoprim-sulfamethoxazole; R, resistance; S, susceptible; +, positive; -, negative.

ESBLs and AmpC β -lactamase genes was found in ten isolates (Table 2).

Four *E. coli* isolates were carrying both *bla*_{TEM} and *bla*_{CTX-M-1} ESBLs genes, and all were positive for *bla*_{CMY-2}. Two of four ESBL producing *K. pneumoniae* was carrying *bla*_{TEM}, *bla*_{SHV} and *bla*_{CTX-M-1} genes, another isolate was a producer of ESBL types SHV and CTX-M-1 subgroup, and one of the four isolates (5125) carrying *bla*_{TEM} also contained the *bla*_{SHV} gene, this being a single *K. pneumoniae* isolate positive for *bla*_{CMY-2} (Table 2).

A significant association of ESBL expression in *E. coli* was observed with resistance to tobramycin ($p \leq 0.001$), tetracycline ($p = 0.043$), and ciprofloxacin ($p \leq 0.001$) (Table 3). A positive test result in *K. pneumoniae* isolates was significantly associated with resistance to tobramycin and ciprofloxacin ($p = 0.006$), and trimethoprim-sulfamethoxazole

($p = 0.043$) (Table 3). Multivariate analyses did not show any association between ESBL production in *E. coli* and *K. pneumoniae*, and resistance to non- β -lactams drugs (Table 4).

Discussion

Community-acquired UTIs caused by ESBL-producing organisms are a cause of concern due to the clinical failure of empirical treatment protocols. This study shows two main pathogens causing UTIs in the community (*E. coli* and *K. pneumoniae*). The results show that expressed resistance to quinolones (ciprofloxacin) is similar among community isolates of *E. coli* and *K. pneumoniae*, 33% and 38%, respectively (Table 1). However, significant resistance was presented to

Table 3 – Crude association between antimicrobial drug resistance and positive test result for ESBL in *E. coli* and *K. pneumoniae*.

Antimicrobial agent	<i>E. coli</i> (n = 102), OR (95% CI)	p	<i>K. pneumoniae</i> (n = 21), OR (95% CI)	p
Amikacin	0.65 (0.07-5.56)	0.932	16 (0.95-267)	0.140
Tobramycin	10.85 (2.87-40.99)	<0.001	55.8 (2.24-1,386.31)	0.006
Tetracycline	8.41(1.04-67.94)	0.043	1.55 (0.11-20.61)	0.710
Ciprofloxacin	57.7 (3.29-1,009.91)	<0.001	55.8 (2.24-1,386.31)	0.006
SXT	3.42 (0.87-13.50)	0.124	22.5 (1.50-335.33)	0.043

OR, odds ratio; CI, confidence interval; SXT, trimethoprim-sulfamethoxazole.
Variables with $p < 0.05$ were considered significant.

Table 4 – Association of antimicrobial drug resistance and presence of ESBL in *E. coli* and *K. pneumoniae*.

Antimicrobial drug	Estimate	OR (95% CI)	p*
<i>E. coli</i>			
Tetracycline	-0.23	0.0002 (0.0000-1.2730)	0.812
Ciprofloxacin	-0.30	0.0001 (0.0000-5.3914)	0.762
Tobramycin	-1.55	0.283 (0.058-1.383)	0.119
SXT	0.19	1.230 (0.159-9.509)	0.842
<i>K. pneumoniae</i>			
Tetracycline	0.54	7.877 (0.004-12678.32)	0.583
Ciprofloxacin	-1.64	0.009 (0.00004-2.4168)	0.100
Tobramycin	-1.24	0.021 (0.00005-0.0407)	0.212
SXT	-0.54	0.119 (0.00005-253.05)	0.586

SXT, trimethoprim-sulfamethoxazole; OR, odds-ratio; CI, confidence interval.

* by χ^2 approximation.

trimethoprim-sulfamethoxazole (50%) and tetracycline (60%) in *E. coli* isolates, characterizing an alarming resistance to these two families of antibiotics.

ESBL production found among community isolates of *E. coli* was 11.7% and 19% for *K. pneumoniae*, rates similar to that found in hospital isolates;^{1,17} *K. pneumoniae* is the main ESBL-producing organism worldwide.¹⁸ In Latin American countries, community-acquired ESBLs are starting to appear with rates up to 28.7% among *E. coli* and *K. pneumoniae* isolates,¹⁹ while in Europe reported rates are of only 1.5% to 5.6%, respectively.²⁰ The presence of ESBL-producing urinary isolates in the community is a significant public health problem that unavoidably increases health care costs for patients and health care providers due to the use of carbapenems, since the oral agents (nitrofurantoin and fosfomicin), used for treatment of UTIs, are unsuitable due to the enormous resistance-development ability demonstrated by uropathogens, which has led to a loss of interest in these drugs. Therefore, carbapenems appear to be the drugs of choice for the treatment of UTIs caused by ESBL-producing *E. coli* and *K. pneumoniae*, considering their rational use.^{21,22}

In Colombia, the prevalence of community-acquired ESBL-producing *E. coli* and *K. pneumoniae* is not well known; the findings of TEM and SHV enzymes in this study show the spread of these enzymes in hospital and community isolates in Colombia.²³ However, the presence of community-acquired CTX-M-producing *Enterobacteriaceae* (seven *E. coli* and three *K. pneumoniae*), is of concern especially in *E. coli*, because these enzymes are commonly encoded by plasmids. Besides, there is the possibility that isolates found in this study are associated with well-characterized ST131 and

ST405, which are multidrug-resistant intercontinental clones responsible for the dissemination of *bla*_{CTX-M-15}, as previously reported in Colombia.²⁴ Multidrug-resistance expressed by ESBL-producing *Enterobacteriaceae* in this study was presented to aminoglycosides, trimethoprim-sulfamethoxazole, tetracycline, and quinolones (Table 2).

ESBL-producing *K. pneumoniae* strains are more related to hospital outbreaks. However, the current distribution of ESBL carrying microorganisms in patients with community-acquired UTIs is a generalized problem, thus producing a flow of isolates from the community to the hospital setting.²⁵ Likewise, the strains of *E. coli* and *K. pneumoniae* that harbored multiple ESBL genes are important, because it has been reported that isolates carrying multiple ESBL genes had a greater resistance to various classes of antibiotic mediated through acquisition of genes located on mobile genetic elements such as plasmids, transposons, or integrons.^{5,26} Furthermore, the fact that they harbor multiple ESBL genes in combination with the gene encoding cephalosporinase CMY-2 could explain the ceftoxitin resistance (four *E. coli* and two *K. pneumoniae*), as reported in previous studies that have shown the *bla*_{CMY-2} gene linked downstream with the promoter sequence ISECP1,²⁷ increasing the likelihood of therapeutic failures and difficulty for laboratory phenotypic detection of ESBL.^{28,29} Finally, resistance expressed by *E. coli* and *K. pneumoniae* isolates, and molecular confirmation of ESBL genes were analyzed using statistical models to measure the crude association between a positive result for ESBL in isolates of *E. coli* and resistance to tobramycin ($p \leq 0.001$), tetracycline ($p = 0.025$) and ciprofloxacin ($p = 0.01$) (Table 3). In *K. pneumoniae* isolates, an

association was found between a positive result for ESBL and resistance to tobramycin ($p=0.002$), ciprofloxacin ($p=0.002$), and trimethoprim-sulfamethoxazole ($p=0.027$) (Table 3). However, in multivariate analysis did not show non- β -lactam drugs as an independent variable. This finding suggests that non- β -lactams resistance remained independently associated with ESBL, but for reasons unrelated to ESBL genes. Studies have shown that non- β -lactam and β -lactam (ESBL) resistance encoding genes are located in plasmids.^{13,30} This common location may explain the resistance to previously mentioned antibiotics. Nevertheless, it is interesting that resistance to ciprofloxacin was associated with presence of ESBL and resistance to other antibiotic classes (Table 2). To some extent, this could be explained by its extensive use as an empirical drug for treatment of UTIs, resulting in strong selective antimicrobial pressure in the community. Similar data were found in previous studies.³¹⁻³³

Conclusion

Detection of CTX-M ESBL in uropathogens isolated from the community is of concern because of the enormous potential for multidrug resistance of strains that produce these enzymes, especially if *E. coli* isolates from this study could be related to the intercontinental clones ST131 and ST405, because empirically administered therapies will fail, which could lead to complicated UTIs.

Conflicts of interest

All authors declare to have no conflict of interest.

Acknowledgements

This work was supported by the Universidad de Córdoba, Institute for Tropical Biological Research.

REFERENCES

- Villegas MV, Correa A, Perez F, et al. Prevalence and characterization of extended-spectrum β -lactamases in *Klebsiella pneumoniae* and *Escherichia coli* isolates from Colombian hospitals. *Diag Microbiol Infect Dis*. 2004;49:217-22.
- Celik AD, Yulugkural Z, Kuloglu F, et al. CTX-M type extended spectrum β -lactamases in *Escherichia coli* isolates from community acquired upper urinary tract infections at a university in the European part of Turkey. *J Microbiol Immunol Infect*. 2010;43:163-7.
- Ambler RP. The structure of beta-lactamases. *Phil Trans R Soc B*. 1980;289:321-31.
- Bonnet R. Growing group of extended-spectrum β -lactamases: the CTX-M enzymes. *Antimicrob Agents Chemother*. 2004;48:1-14.
- Apisarnthanarak A, Kiratisin P, Mundy L. Clinical and molecular epidemiology of healthcare-associated infections due to extended-spectrum β -lactamase (ESBL)-producing strains of *Escherichia coli* and *Klebsiella pneumoniae* that harbor multiple ESBL genes. *Infect Control Hosp Epidemiol*. 2008;29:1026-34.
- Nicolas-Chanoine M, Blanco J, Leflon-Guibout V, et al. Intercontinental emergence of *Escherichia coli* clone O25:H4-ST131 producing CTX-M-15. *J Antimicrob Chemother*. 2008;61:273-81.
- Ben-Ami R, Schwaber MJ, Navon-Venezia S, et al. Influx of extended-spectrum β -lactamase-producing Enterobacteriaceae into the hospital. *Clin Infect Dis*. 2006;42:925-34.
- Guidoni E, Berezin E, Nigro S, et al. Antibiotic resistance patterns of pediatric community-acquired urinary infections. *Braz J Infect Dis*. 2008;12:321-3.
- Farmer J. Enterobacteriaceae: introduction and identification. In: Murray P, Baron E, Pfaller M, Tenover F, Tenover F, editors. *Manual of clinical microbiology*. 7th ed. Washington DC: American Society of Microbiology; 1999. p. 442-58.
- Clinical and Laboratory Standards Institute. Performance standards for antimicrobial susceptibility testing; fourteenth informational supplement. CLSI document M100-S21. Wayne, PA: Clinical and Laboratory Standards Institute (CLSI);2011.
- Carter MW, Oakton KJ, Warner M, Livermore DM. Detection of extended-spectrum β -lactamases in *Klebsiellae* with the oxoid combination disk method. *J Clin Microbiol*. 2000;38:4228-32.
- Dubois V, Arpin C, Noury P, et al. Prolonged outbreak of infection due to TEM-21-producing strains of *Pseudomonas aeruginosa* and Enterobacteria in a nursing home. *J Clin Microbiol*. 2005;43:4129-38.
- Silva J, Gatica R, Aguilar C, et al. Outbreak of infection with extended-spectrum β -lactamase-producing *Klebsiella pneumoniae* in a Mexican hospital. *J Clin Microbiol*. 2001;39:3193-6.
- Villegas MV, Correa A, Perez F, et al. CTX-M-12 β -lactamase in a *Klebsiella pneumoniae* clinical isolate in Colombia. *Antimicrob Agents Chemother*. 2004;48:629-31.
- Paterson DL, Hujer KM, Hujer AM, et al. Extended-spectrum β -lactamases in *Klebsiella pneumoniae* bloodstream isolates from seven countries: dominance and widespread prevalence of SHV- and CTX-M-type β -lactamases. *Antimicrob Agents Chemother*. 2003;47:3554-60.
- Bauernfeind A, Stemplinger I, Jungwirth R, et al. Characterization of beta-lactamase gene bla_{PER-2}, which encodes an extended-spectrum class A beta-lactamase. *Antimicrob Agents Chemother*. 1996;40:616-20.
- Martinez P, Espinal P, Máttar S. Epidemiología molecular de *Pseudomonas aeruginosa* resistente a β -lactámicos de amplio espectro en el Hospital San Jerónimo de Montería. *Infectio*. 2007;11:6-15.
- Demirdag K, Hosoglu S. Epidemiology and risk factors for ESBL-producing *Klebsiella pneumoniae*: a case control study. *J Infect Dev Ctries*. 2010;4:717-22.
- Villegas MV, Blanco MG, Sifuentes-Osornio J, Rossi F. Increasing prevalence of extended-spectrum-beta-lactamase among Gram-negative bacilli in Latin America: 2008 update from the Study for Monitoring Antimicrobial Resistance Trends (SMART). *Braz J Infect Dis*. 2011;15:34-9.
- Schito GC, Naber KG, Botto H, et al. The ARESC study: an international survey on the antimicrobial resistance of pathogens involved in uncomplicated urinary tract infections. *Inter J Antimicrob Agents*. 2009;34:407-13.
- Falagas ME, Grammatikos AP, Michalopoulos A. Potential of old-generation antibiotics to address current need for new antibiotics. *Expert Rev Anti Infect Ther*. 2008;6:593-600.
- Pitout J, Laupland K. Extended-spectrum β -lactamase-producing Enterobacteriaceae: an emerging public-health concern. *Lancet Infect Dis*. 2008;8:159-66.
- Villegas MV, Kattan JN, Quinteros MG, Casellas JM. Prevalence of extended-spectrum beta-lactamases in South America. *Clin Microbiol Infect*. 2008;14 Suppl. 1:154-8.

24. Ruiz SJ, Montealegre MC, Ruiz-Garbajosa P, et al. First characterization of CTX-M-15-producing *Escherichia coli* ST131 and ST405 clones causing community-onset infections in South America. *J Clin Microbiol*. 2011;49:1993-6.
25. Keynan Y, Rubinstein E. The changing face of *Klebsiella pneumoniae* infections in the community. *Inter J Antimicrob Agents*. 2007;30:385-9.
26. Motta RN, Oliveira MM, Magalhães PS, et al. Plasmid-mediated extended-spectrum β -lactamase-producing strains of Enterobacteriaceae isolated from diabetes foot infections in a Brazilian diabetic center. *Braz J Infect Dis*. 2003;7:129-34.
27. Haldorsen B, Aasnaes B, Dahl K, et al. The AmpC phenotype in Norwegian clinical isolates of *Escherichia coli* is associated with an acquired ISEcp1-like ampC element or hyperproduction of the endogenous AmpC. *J Antimicrob Chemother*. 2008;62:694-702.
28. Navarro F, Calvo J, Cantón R, Fernández-Cuenca F, Mirelis B. Detección fenotípica de mecanismos de resistencia en microorganismos gramnegativos. *Enferm Infecc Microbiol Clín*. 2011;29:524-34.
29. Martínez P, Máttar S. Emergencia de la resistencia antibiótica debida a las β -lactamasas de espectro extendido (BLEE): detección, impacto clínico y epidemiología. *Infectio*. 2007;11:23-35.
30. Leverstein-van Hall MA, M.Blok HE, Donders AR, Paauw A, Fluit AC, Verhoef J. Multidrug resistance among Enterobacteriaceae is strongly associated with the presence of integrons and is independent of species or isolate origin. *J Infect Dis*. 2003;187:251-9.
31. Paterson DL, Mulazimoglu L, Casellas JM, et al. Epidemiology of ciprofloxacin resistance and its relationship to extended-spectrum β -lactamase production in *Klebsiella pneumoniae* isolates causing bacteremia. *Clin Infect Dis*. 2000;30:473-8.
32. Akpaka PE, Swanston WH. Phenotypic detection and occurrence of extended-spectrum beta-lactamases in clinical isolates of *Klebsiella pneumoniae* and *Escherichia coli* at a tertiary hospital in Trinidad & Tobago. *Braz J Infect Dis*. 2008;12:516-20.
33. Serefhanoglu K, Turan H, Timurkaynak F, Arslan H. Bloodstream infections caused by ESBL-producing *E. coli* and *K. pneumoniae*: risk factors for multidrug-resistance. *Braz J Infect Dis*. 2009;13:403-7.